

# Fast and selective synthesis of novel cyclic sulfamide HIV-1 protease inhibitors under controlled microwave heating

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**Abstract**—A novel and highly selective silver-promoted monobenylation method was developed to promote synthesis of nonsymmetrical sulfamide-based HIV-1 inhibitors. Microwave-accelerated palladium-catalyzed *N*-amide arylation- and aminocarbonylation reactions were employed for rapid and reliable compound generation. With this class of inhibitory agents, six active inhibitors were identified, the most potent inhibitor possessing a  $K_i$ -value of 20 nM.

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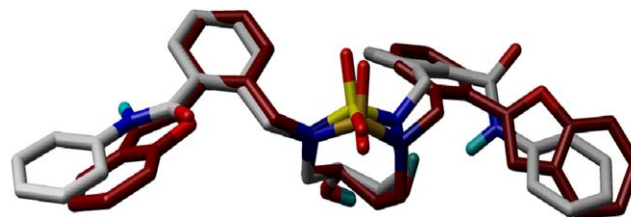
## 1. Introduction

Since the mid-1980s, a large number of studies have demonstrated that an acceleration of chemical rates can be achieved by employing high-density microwave irradiation instead of traditional sources of heat.<sup>1–4</sup> Although high yields and clean reactions are commonly obtained with microwave heating, reduced selectivities have been reported at high temperatures.<sup>5,6</sup> Hence, development of fast and highly selective reaction protocols, remains a challenge.

The recent development of palladium-catalyzed gas-free aminocarbonylations<sup>7</sup> and *N*-amide arylations<sup>8</sup> has enabled direct attachment of amide functionalities to  $sp^2$ -carbons that were previously difficult to accomplish. However, the commonly tedious fine-tuning of the appropriate reaction parameters, and the requirement for inert conditions and long reaction times, has limited the usage of these direct transformations in medicinal chemistry.

A large number of very potent urea-based cyclic HIV-1 protease inhibitors carrying four-side chains has been prepared and evaluated following the pioneering work by Lam and co-workers.<sup>9</sup> Interestingly, by switching the water-mimicking group from urea to sulfamide, an unanticipated flipped binding mode was obtained according to X-ray crystal structures

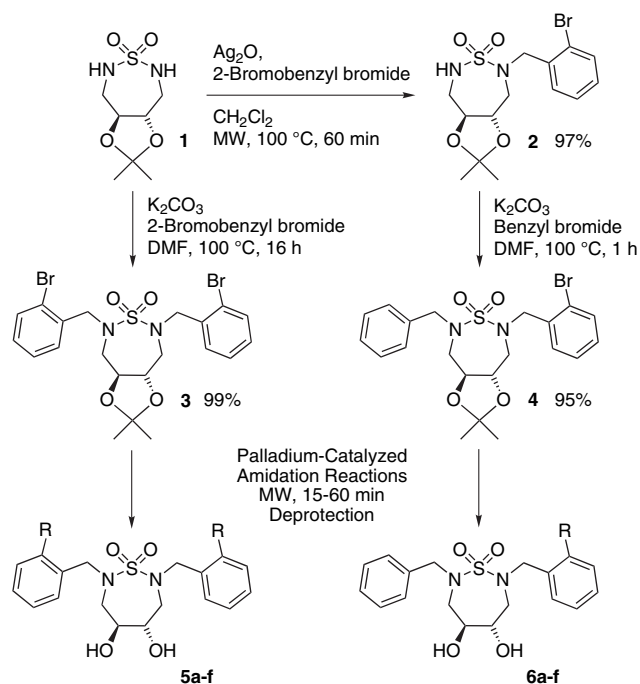
of inhibitors in complex with the HIV-1 protease.<sup>10,11</sup> The preparative route to these inhibitors was complex and the inhibitors were lipophilic and of high-molecular weight. To investigate if properly functionalized benzylic side-chains could span from *P2/P2'* to *P1/P1'* and thus simplify the otherwise cumbersome synthetic pathway, a set of  $C_2$ -symmetric *ortho*-functionalized sulfamide derivatives were synthesized and evaluated. The most potent inhibitor from this series ( $K_i=0.53 \mu\text{M}$ ), substituted with two benzofuran moieties, was identified as a lead structure for further optimization (Fig. 1).<sup>12</sup> Thus, based on isosteric replacement and modeling, it was hypothesized that an amide function ( $-\text{CONH}$ ) might be of interest to incorporate in the two *ortho*-positions (*R*-) of the dibenzylated cyclic sulfamide **5** to act as a mimic for the furan ring (Fig. 1, Scheme 1). In addition, we decided to investigate the inverted amides ( $-\text{NHCO}$ ). Reactants were selected in order to vary flexibility and size of the *ortho*-substituents. With the aim to reduce



**Figure 1.** Superposition of benzofuran and anilide (**5a**) *o*-substituted  $N,N'$ -dibenzylcyclic seven-membered sulfamide structures. The molecular graphics image was produced using the UCSF program Chimera.<sup>13</sup>

**Keywords:** Microwave; Aminocarbonylation; Goldberg; HIV-1 protease inhibitor.

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Scheme 1.

the size of the inhibitors and to produce nonsymmetrically decorated dibenzyl sulfamides, precursor **2** was much desired.

In this communication, a microwave promoted and chemoselective procedure for monoalkylation of the sulfamide scaffold **1** is reported (Scheme 1). Furthermore, a number of direct palladium-catalyzed *ortho*-amidations of the *P2/P2'* benzyl groups have been conducted, delivering 12 new HIV-1 protease inhibitors.

## 2. Results and discussion

### 2.1. Synthetic strategies

The core structure **1** served as the precursor to provide the symmetric and nonsymmetric aryl bromide derivatives **3** and **4**, respectively (Scheme 1). Compound **3** was obtained, as previously reported, in 99% yield after *N,N*-dibenylation with 2-bromobenzyl bromide in the presence of  $K_2CO_3$ .<sup>10</sup> Double *ortho*-amidations of **3** by palladium-catalyzed coupling reactions and deprotection, provided the symmetrical inhibitors in good yields (Table 1). More specifically, the aminocarbonylation products **5a,b** were obtained in 59 and 80% yield after 60 min of heating at 150 °C, respectively.<sup>14</sup> The corresponding *N*-amide arylations delivered the inverted amides **5c–f** in 53–72% yields after only 15 min of irradiation (160 °C). Early attempts to prepare unsymmetrical **4** from **1** showed an increased reactivity of the monoalkylated product **2** towards concomitant dialkylation, resulting in unwanted **3**. Fortunately, by initial addition of 1.5 equiv  $Ag_2O$ , subsequent microwave-assisted benzylation afforded pure aryl bromide **2** with excellent selectivity (**2/3**=99:1, Scheme 1). The yield of **2** was 97% despite the high-reaction temperature (100 °C). Related monoalkylations of symmetrical diols have been performed in presence of  $Ag_2O$ .<sup>15</sup>

**Table 1.** Microwave-heated palladium-catalyzed coupling reactions on symmetric and nonsymmetric cyclic sulfamides **3** and **4**

Reactant <sup>a</sup> , route <sup>b</sup>	R-group	Symmetric (yield) <sup>c</sup> , $K_i$ (nM)	Nonsymmetric (yield) <sup>c</sup> , $K_i$ (nM)
a, A		<b>5a</b> (59%), >20,000	<b>6a</b> (77%), >20,000
b, A		<b>5b</b> (80%), 8600	<b>6b</b> (74%), >20,000
c, B		<b>5c</b> (72%), >20,000	<b>6c</b> (77%), >20,000
d, B		<b>5d</b> (53%), 1200	<b>6d</b> (51%), >20,000
e, B		<b>5e</b> (54%), 1300	<b>6e</b> (68%), 7700
f, B		<b>5f</b> (57%), 20	<b>6f</b> (66%), 140

<sup>a</sup> a: Aniline, b: Benzylamine, c: Benzamide, d: 2-Phenylacetamide, e: 2-(3-Methoxyphenyl)acetamide, f: 2-(2-Naphthyl)acetamide.

<sup>b</sup> A: Aminocarbonylation, B: *N*-amide arylation.

<sup>c</sup> Isolated yields after deprotection.

Aminocarbonylations of parent **4** with  $Mo(CO)_6$  as the CO-source smoothly produced the monofunctionalized products **6a,b** (74 and 77%, Table 1). Similarly, **6c–f** were prepared as reported for the symmetric analogs **5c–f** in 51–77% yields after deprotection. The  $K_i$ -values for the synthesized compounds were determined as previously described.<sup>16,17</sup>

### 2.2. Discussion

Six active inhibitors were identified. Apparently, incorporation of the amide function alone was not sufficient, and an extra methylene spacer was required in order to yield active symmetrical compounds (**5b**, **5d–f**). The most potent compound **5f** possessed low nanomolar activity with a  $K_i$ -value of 20 nM. The high activity of **5f** was obtained by steric replacement of phenylacetamide for 1-naphthylacetamide. To our satisfaction, the smaller nonsymmetric compounds, occupying only three subsites, also proved to be active (**6e**, **6f**). With these structures, both the extra methylene spacer and further enlargement of the R-group was essential for activity.

## 3. Conclusion

In conclusion, the concept of *ortho*-extension from benzylic *P2/P2'* side-chains to reach the *P1/P1'* binding sites provided highly active HIV-1 protease inhibitors. With regard to inhibition potency, improved  $K_i$ -values were achieved using mono- and di-*ortho*-elongated structures with flexible three-atom spacers between the aromatic moieties.

Furthermore, single-mode microwave heating at 100 °C for 60 min was exploited without compromising the selectivity in the key monobenylation step.

## 4. Experimental

### 4.1. General

The microwave-assisted reactions were performed in a single-mode microwave cavity (Smith Synthesizer, Biotage AB, Uppsala, Sweden) producing controlled irradiation at 2450 MHz. Reaction temperatures were determined and controlled via the built-in, on-line IR-sensor. Flash column chromatography was performed using Merck Silica gel 60 (0.040–0.063 mm). Analytical HPLC–MS analyses were performed using a Gilson HPLC system with a Chromolith SpeedROD RP-18e column (50×4.6 mm) and a Finnigan AQA quadropole mass spectrometer using a 4 mL/min CH<sub>3</sub>CN/H<sub>2</sub>O gradient (0.05% HCOOH), employing UV-detection (214 and 254 nm) and mass selective detector (ESI+). <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a Varian Mercury-400 spectrometer at 399.8 and 100.6 MHz, respectively. All starting materials and reagents were commercially available and used as received. Xantphos, (4,5-bis(diphenylphosphino)-9,9-dimethyl-xanthene) was purchased from Aldrich. Herrmann's palladacycle, *trans*-di(μ-acetato)-bis[*o*-(di-*o*-tolylphosphino)benzyl]dipalladium (II) was purchased from Strem and Mo(CO)<sub>6</sub> was obtained from Acros.

### 4.2. Method for monobenylation of cyclic sulfamides

Cyclic sulfamide **1** (0.45 mmol, 100 mg) and Ag<sub>2</sub>O (0.67 mmol, 156 mg) were mixed in a Smith process vial for 5 min in 2 mL of CH<sub>2</sub>Cl<sub>2</sub>. After addition of *o*-bromobenzyl bromide (0.47 mmol, 117 mg) and another 3 mL of CH<sub>2</sub>Cl<sub>2</sub>, the vial was capped with a septum. The microwave synthesizer was set to 100 °C for 1 h. After cooling, the reaction mixture was filtered through a plug of Celite and immediately transferred to a short flash column and **2** was easily purified using 9:1 *iso*-hexane/EtOAc as the eluent (>95% purity by <sup>1</sup>H NMR and GC–MS). In a subsequent benzylation, **2** (0.68 mmol, 266 mg), benzyl bromide (1.36 mmol, 232 mg) and K<sub>2</sub>CO<sub>3</sub> (3.33 mmol, 460 mg) were mixed in a process vial with 5 mL DMF. Reaction was heated in a heating block at 60 °C overnight. The reaction mixture was concentrated in vacuo and purified over silica, using 2:1–1:1 *iso*-hexane/CH<sub>2</sub>Cl<sub>2</sub> as the eluent.

**4.2.1. 3,4,5,6-Tetrahydro-(4S,5S)-2-(2-bromobenzyl)-4,5-O-isopropylidene-1,2,7-thiadiazepine 1,1-dioxide (2).** White solid. LC–MS, *m/z*=392 [M+H<sup>+</sup>]. <sup>1</sup>H NMR (400 MHz, acetone-*d*<sub>6</sub>): δ=7.62 (dd, 1H, *J*=1.2, 8.0 Hz), 7.56 (dd, 1H, *J*=1.7, 7.8 Hz), 7.44 (ddd, 1H, *J*=1.2, 7.4, 7.8 Hz), 7.26 (ddd, 1H, *J*=1.7, 7.4, 8.0 Hz), 6.72 (br m, 1H), 4.51 (d, 1H, *J*=16.0 Hz), 4.46 (d, 1H, *J*=16.0 Hz), 4.32–4.22 (m, 2H), 3.59 (ddd, 1H, *J*=3.9, 4.7, 12.8 Hz), 3.50 (dd, 1H, *J*=4.2, 13.6 Hz), 3.15 (ddd, 1H, *J*=0.6, 9.4, 13.6 Hz), 3.08 (ddd, 1H, *J*=7.0, 9.5, 12.8 Hz), 1.37 (m, 3H), 1.36 (m, 3H). <sup>13</sup>C NMR (100.5 MHz, acetone-*d*<sub>6</sub>): δ=136.5, 132.9, 1230.0, 129.6, 128.3, 128.2, 109.2, 78.7, 77.8, 54.0, 49.4, 43.5, 26.6, 26.5. Anal. Calcd (%) for

C<sub>14</sub>H<sub>19</sub>BrN<sub>2</sub>O<sub>4</sub>S: C, 42.98; H, 4.89; N, 7.16. Found: C, 43.30; H, 5.04; N, 7.19.

**4.2.2. 3,4,5,6-Tetrahydro-(4S,5S)-2-(2-bromobenzyl)-7-benzyl-4,5-O-isopropylidene-1,2,7-thiadiazepine 1,1-dioxide (4).** White solid. LC–MS, *m/z*=482 [M+H<sup>+</sup>]. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ=7.57 (m, 1H), 7.54 (m, 1H), 7.40–7.29 (m, 6H), 7.18 (m, 1H), 4.58 (d, 1H, *J*=15.4 Hz), 4.50 (d, 1H, *J*=15.4 Hz), 4.49 (d, 1H, *J*=14.6 Hz), 4.36 (d, 1H, *J*=14.6 Hz), 4.31–4.18 (m, 2H), 3.50 (dd, 1H, *J*=4.5, 13.0 Hz), 3.46 (dd, 1H, *J*=4.6, 12.8 Hz), 3.11 (dd, 1H, *J*=9.2, 13.0 Hz), 3.02 (dd, 1H, *J*=9.3, 12.8 Hz), 1.38 (d, 1H, *J*=0.6 Hz), 1.37 (d, 1H, *J*=0.6 Hz). <sup>13</sup>C NMR (100.5 MHz, CDCl<sub>3</sub>): δ=136.1, 135.5, 133.2, 130.3, 129.7, 129.0, 128.7, 128.4, 128.2, 123.8, 110.1, 77.9, 77.7, 55.4, 54.8, 49.3, 48.6, 27.1. Anal. Calcd (%) for C<sub>21</sub>H<sub>25</sub>N<sub>2</sub>O<sub>4</sub>S: C, 52.39; H, 5.23; N, 5.82. Found: C, 52.46; H, 5.29; N, 5.79.

### 4.3. General method for aminocarbonylation of both symmetrical (3) and nonsymmetrical (4) sulfamide scaffolds

Cyclic sulfamide (0.083 mmol), Herrmann's catalyst (8.3 μmol, 7.8 mg), Mo(CO)<sub>6</sub> (0.017 mmol, 47 mg), DBU (0.83 mmol, 136 mg) and amine (2.5 mmol) were put in a 2–5 mL Smith process vial. THF (2.5 mL) was added and the reaction was run at 150 °C for 60 min. After cooling the reaction mixture was filtered through a plug of Celite and the organic solvent was evaporated under reduced pressure. Flash column chromatography and removal of the protective group using 1 mL 2.2 M HCl/ether and 2 mL methanol at rt for 45 min, followed by a second rapid flash column chromatography yielded pure products in all cases.

**4.3.1. 3,4,5,6-Tetrahydro-(4S,5S)-2,7-bis(*N*-phenyl-2-amido-benzyl)-4,5-dihydroxy-1,2,7-thiadiazepine 1,1-dioxide (5a).** White solid. LC–MS, *m/z*=601 [M+H<sup>+</sup>]. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ=8.09 (s, 2H), 7.62–7.54 (m, 6H), 7.46–7.40 (m, 4H), 7.36–7.24 (m, 6H), 7.15 (tt, 2H, *J*=7.4, 1.2 Hz), 4.64 (d, 2H, *J*=15.8 Hz), 4.58 (d, 2H, *J*=15.8 Hz), 3.61–3.56 (m, 2H), 3.35 (d, 1H, *J*=15.4 Hz), 3.34 (d, 1H, *J*=15.4 Hz), 3.22 (d, 1H, *J*=15.4 Hz), 3.20 (d, 1H, *J*=15.4 Hz). <sup>13</sup>C NMR (100.5 MHz, CDCl<sub>3</sub>): δ=168.3, 137.9, 136.2, 136.0, 131.3, 130.2, 129.4, 128.2, 127.1, 125.2, 120.5, 72.4, 50.6, 48.9.

**4.3.2. 3,4,5,6-Tetrahydro-(4S,5S)-2,7-bis(*N*-benzyl-2-amido-benzyl)-4,5-dihydroxy-1,2,7-thiadiazepine 1,1-dioxide (5b).** White solid. LC–MS, *m/z*=629 [M+H<sup>+</sup>]. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ=7.68 (ddd, 2H, *J*=7.8, 1.2, 0.6 Hz), 7.49 (ddd, 2H, *J*=7.8, 7.3, 1.5 Hz), 7.42–7.27 (m, 14H), 6.34 (br t, 2H, *J*=5.8 Hz), 4.78 (d, 2H, *J*=15.4 Hz), 4.73 (d, 2H, *J*=15.4 Hz), 4.64 (dd, 2H, *J*=14.7, 5.8 Hz), 4.59 (dd, 2H, *J*=14.7, 5.8 Hz), 3.76–3.72 (m, 2H), 3.58–3.54 (m, 2H), 3.53–3.47 (m, 2H), 3.27–3.20 (m, 2H). <sup>13</sup>C NMR (100.5 MHz, CDCl<sub>3</sub>): δ=169.9, 137.8, 136.2, 135.8, 131.3, 130.9, 129.2, 128.22, 128.18, 128.1, 126.9, 72.5, 49.9, 47.2, 44.5.

**4.3.3. 3,4,5,6-Tetrahydro-(4S,5S)-2-(*N*-phenyl-2-amido-benzyl)-7-benzyl-4,5-dihydroxy-1,2,7-thiadiazepine 1,1-dioxide (6a).** White solid. LC–MS, *m/z*=482 [M+H<sup>+</sup>]. <sup>1</sup>H NMR (400 MHz, acetone-*d*<sub>6</sub>): δ=9.57 (br s, 1H),

7.87–7.82 (m, 2H), 7.70 (m, 1H), 7.62 (m, 1H), 7.55 (m, 1H), 7.46–7.27 (m, 8H), 7.13 (m, 1H), 4.97 (d, 1H,  $J=16.8$  Hz), 4.82 (d, 1H,  $J=16.8$  Hz), 4.68 (d, 1H,  $J=15.8$  Hz), 4.55 (d, 1H,  $J=15.8$  Hz), 4.26 (d, 1H,  $J=3.9$  Hz), 4.22 (d, 1H,  $J=4.3$  Hz), 3.42–3.23 (m, 3H), 3.15 (dd, 1H,  $J=15.2$ , 3.2 Hz).  $^{13}\text{C}$  NMR (100.5 MHz, acetone- $d_6$ ):  $\delta=168.2$ , 140.3, 138.5, 137.3, 137.3, 131.2, 129.6, 129.4, 129.2, 128.8, 128.34, 128.30, 128.0, 124.7, 120.8, 73.1, 73.0, 53.2, 50.9, 49.3, 48.5. Anal. Calcd (%) for  $\text{C}_{25}\text{H}_{27}\text{N}_3\text{O}_5\text{S}$ : C, 62.35; H, 5.65; N, 8.73. Found: C, 62.21; H, 5.80; N, 8.61.

**4.3.4. 3,4,5,6-Tetrahydro-(4*S*,5*S*)-2-(*N*-benzyl-2-amido-benzyl)-7-benzyl-4,5-dihydroxy-1,2,7-thiadiazepine 1,1-dioxide (6b).** White solid. LC–MS,  $m/z=496$  [ $\text{M}+\text{H}^+$ ].  $^1\text{H}$  NMR (400 MHz, acetone- $d_6$ ):  $\delta=8.13$  (br t, 1H), 7.67 (ddd, 1H,  $J=7.8$ , 1.2, 0.6 Hz), 7.57–7.22 (m, 13H), 4.89 (d, 1H,  $J=16.6$  Hz), 4.78 (d, 1H,  $J=16.6$  Hz), 4.70 (d, 1H,  $J=15.8$  Hz), 4.61 (d, 2H,  $J=6.0$  Hz), 4.56 (d, 1H,  $J=15.8$  Hz), 4.29 (d, 1H,  $J=3.8$  Hz), 4.26 (d, 1H,  $J=4.2$  Hz), 3.67–3.56 (m, 2H), 3.40–3.12 (m, 4H).  $^{13}\text{C}$  NMR (100.5 MHz, acetone- $d_6$ ):  $\delta=169.7$ , 140.3, 138.6, 137.2, 137.1, 131.0, 129.42, 129.40, 129.3, 128.9, 128.4, 128.3, 128.1, 128.1, 127.8, 73.1, 73.0, 53.2, 50.6, 49.0, 48.5, 43.9. Anal. Calcd (%) for  $\text{C}_{26}\text{H}_{29}\text{N}_3\text{O}_5\text{S}$ : C, 63.01; H, 5.90; N, 8.48. Found: C, 62.89; H, 6.06; N, 8.43.

#### 4.4. General method for *N*-amide arylation of both symmetrical (3) and nonsymmetrical (4) sulfamide scaffolds

Cyclic sulfamide (0.083 mmol),  $\text{Pd}(\text{dba})_2$  (4  $\mu\text{mol}$ , 2.4 mg), Xantphos (6.2  $\mu\text{mol}$ , 3.6 mg),  $\text{Cs}_2\text{CO}_3$  (0.4 mmol, 135 mg) and amide (0.8 mmol) were put in a Smith process vial, with 2.5 mL 10% NMP in dioxane as the solvent. The reaction mixture was heated in the microwave at 160 °C for 15 min. After cooling the reaction mixture was filtered through a plug of Celite and the organic solvent was evaporated under reduced pressure. Flash column chromatography and removal of the protecting group using 1 mL of 2.2 M HCl/ether and 2 mL methanol at rt for 45 min followed by a second rapid flash column chromatography yielded pure products in all cases.

**4.4.1. 3,4,5,6-Tetrahydro-(4*S*,5*S*)-2,7-bis(2-benzamido-benzyl)-4,5-dihydroxy-1,2,7-thiadiazepine 1,1-dioxide (5c).** White solid. LC–MS,  $m/z=601$  [ $\text{M}+\text{H}^+$ ].  $^1\text{H}$  NMR (400 MHz, acetone- $d_6$ ):  $\delta=9.31$  (br s, 2H), 8.05 (dd, 4H,  $J=1.4$ , 8.4 Hz), 7.80 (dd, 2H,  $J=1.4$ , 8.0 Hz), 7.61–7.56 (m, 2H), 7.55–7.47 (m, 6H), 7.38 (ddd, 2H,  $J=1.6$ , 7.5, 7.7 Hz), 7.26 (ddd, 2H,  $J=1.4$ , 7.5, 7.5 Hz), 4.72 (d, 2H,  $J=16.2$  Hz), 4.68 (d, 2H,  $J=16.2$  Hz), 4.32 (br d, 2H,  $J=4.1$  Hz), 3.54–3.45 (m, 2H), 3.30 (dd, 2H,  $J=9.2$ , 15.1 Hz), 3.18 (dd, 2H,  $J=2.9$ , 15.1 Hz).  $^{13}\text{C}$  NMR (100.5 MHz, acetone- $d_6$ ):  $\delta=166.15$ , 137.0, 135.0, 131.9, 130.6, 129.7, 128.6, 128.5, 127.9, 125.9, 125.7, 72.6, 49.5, 48.3.

**4.4.2. 3,4,5,6-Tetrahydro-(4*S*,5*S*)-2,7-bis[2-(2-phenylacetamido)-benzyl]-4,5-dihydroxy-1,2,7-thiadiazepine 1,1-dioxide (5d).** White solid. LC–MS,  $m/z=629$  [ $\text{M}+\text{H}^+$ ].  $^1\text{H}$  NMR (400 MHz, acetone- $d_6$ ):  $\delta=8.92$  (s, 2H), 7.94 (dm, 2H,  $J=8.1$  Hz), 7.49 (dm, 2H,  $J=7.5$  Hz), 7.42 (dm, 4H,  $J=7.5$  Hz), 7.37–7.26 (m, 6H), 7.25–7.15 (m, 4H),

4.68 (d, 2H,  $J=15.5$  Hz), 4.52 (d, 2H,  $J=15.5$  Hz), 4.23 (br s, 2H), 3.76 (s, 4H), 3.30–3.17 (m, 4H), 3.16–3.02 (m, 2H).  $^{13}\text{C}$  NMR (100.5 MHz, acetone- $d_6$ ):  $\delta=169.6$ , 137.5, 136.2, 130.1, 129.5, 128.9, 128.6, 128.0, 126.9, 124.9, 123.7, 72.5, 49.5, 48.5, 43.9.

**4.4.3. 3,4,5,6-Tetrahydro-(4*S*,5*S*)-2,7-bis[2-[2-(3-methoxy-phenyl)-acetamido]-benzyl]-4,5-dihydroxy-1,2,7-thiadiazepine 1,1-dioxide (5e).** White solid. LC–MS,  $m/z=689$  [ $\text{M}+\text{H}^+$ ].  $^1\text{H}$  NMR (400 MHz, acetone- $d_6$ ):  $\delta=8.88$  (s, 2H), 7.93 (dm, 2H,  $J=8.1$  Hz), 7.48 (dd, 2H,  $J=1.6$ , 7.6 Hz), 7.34 (ddd, 2H,  $J=1.6$ , 7.6, 7.6 Hz), 7.23–7.15 (m, 4H), 7.02 (m, 2H), 6.99 (dm, 2H,  $J=7.6$  Hz), 6.80 (ddd, 2H,  $J=1.0$ , 2.6, 8.2 Hz), 4.68 (d, 2H,  $J=15.5$  Hz), 4.52 (d, 2H,  $J=15.5$  Hz), 4.21 (br s, 2H), 3.77 (s, 6H), 3.73 (s, 4H), 3.32–3.18 (m, 4H), 3.17–3.08 (m, 2H).  $^{13}\text{C}$  NMR (100.5 MHz, acetone- $d_6$ ):  $\delta=167.8$ , 146.2, 138.7, 138.2, 137.9, 130.1, 129.8, 129.7, 129.3, 128.8, 126.9, 73.1, 50.9, 49.0, 46.2, 44.1.

**4.4.4. 3,4,5,6-Tetrahydro-(4*S*,5*S*)-2,7-bis[2-[2-(2-naphthyl)-acetamido]-benzyl]-4,5-dihydroxy-1,2,7-thiadiazepine 1,1-dioxide (5f).** White solid. LC–MS,  $m/z=729$  [ $\text{M}+\text{H}^+$ ].  $^1\text{H}$  NMR (400 MHz, acetone- $d_6$ ):  $\delta=8.91$  (s, 2H), 8.25 (ddd, 2H,  $J=0.9$ , 2.1, 8.5 Hz), 7.90 (dm, 2H,  $J=8.2$  Hz), 7.83 (dd, 2H,  $J=1.2$ , 8.0 Hz), 7.82 (dm, 2H,  $J=8.2$  Hz), 7.60–7.39 (m, 10H), 7.31 (ddd, 2H,  $J=1.6$ , 7.5, 7.7 Hz), 7.16 (ddd, 2H,  $J=1.3$ , 7.5, 7.5 Hz), 4.63 (d, 2H,  $J=15.6$  Hz), 4.51 (d, 2H,  $J=15.6$  Hz), 4.27 (s, 4H), 4.22 (br s, 2H), 3.32–3.22 (m, 4H), 3.20–3.08 (m, 2H).  $^{13}\text{C}$  NMR (100.5 MHz, acetone- $d_6$ ):  $\delta=170.2$ , 138.0, 134.9, 133.4, 133.1, 130.6, 129.43, 129.39, 129.3, 129.0, 128.5, 127.1, 126.6, 126.4, 125.8, 125.3, 124.9, 73.2, 50.1, 49.2, 42.1. Anal. Calcd (%) for  $\text{C}_{42}\text{H}_{40}\text{N}_4\text{O}_6\text{S}$ : C, 69.2; H, 5.53; N, 7.69. Found: C, 68.95; H, 5.66; N, 7.50.

**4.4.5. 3,4,5,6-Tetrahydro-(4*S*,5*S*)-2-(2-benzamidobenzyl)-7-benzyl-4,5-dihydroxy-1,2,7-thiadiazepine 1,1-dioxide (6c).** White solid. LC–MS,  $m/z=482$  [ $\text{M}+\text{H}^+$ ].  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta=8.73$  (br s, 1H), 7.93 (dd, 2H,  $J=1.3$ , 8.4 Hz), 7.86 (dm, 1H,  $J=8.1$  Hz), 7.55 (m, 2H), 7.49–7.42 (m, 2H), 7.39–7.23 (m, 7H), 7.20 (m, 1H), 4.60 (d, 1H,  $J=14.8$  Hz), 4.58 (d, 1H,  $J=15.5$  Hz), 4.34 (d, 1H,  $J=14.8$  Hz), 4.30 (d, 1H,  $J=15.5$  Hz), 3.42 (m, 1H), 3.30 (dd, 1H,  $J=9.2$ , 14.6 Hz), 3.26–3.08 (m, 4H), 3.03 (dd, 1H,  $J=3.9$ , 15.2 Hz), 2.84 (br s, 1H).  $^{13}\text{C}$  NMR (100.5 MHz,  $\text{CDCl}_3$ ):  $\delta=167.7$ , 136.5, 136.2, 134.0, 132.5, 131.1, 129.7, 129.1, 128.9, 128.44, 128.43, 128.2, 127.9, 126.4, 125.7, 73.2, 72.4, 53.2, 50.0, 48.7, 47.9. Anal. Calcd (%) for  $\text{C}_{25}\text{H}_{27}\text{N}_3\text{O}_5\text{S}$ : C, 62.35; H, 5.65; N, 8.73. Found: C, 62.51; H, 5.79; N, 8.64.

**4.4.6. 3,4,5,6-Tetrahydro-(4*S*,5*S*)-2-(2-phenylacetamido-benzyl)-7-benzyl-4,5-dihydroxy-1,2,7-thiadiazepine 1,1-dioxide (6d).** White solid. LC–MS,  $m/z=496$  [ $\text{M}+\text{H}^+$ ].  $^1\text{H}$  NMR (400 MHz, acetone- $d_6$ ):  $\delta=8.93$  (s, 1H), 8.06 (dd, 1H,  $J=1.2$ , 8.1 Hz), 7.51–7.39 (m, 7H), 7.37–7.30 (m, 2H), 7.29–7.18 (m, 3H), 7.14 (ddd, 1H,  $J=1.3$ , 7.5, 7.5 Hz), 4.78 (d, 1H,  $J=15.7$  Hz), 4.70 (d, 1H,  $J=15.3$  Hz), 4.54 (d, 1H,  $J=15.7$  Hz), 4.42 (d, 1H,  $J=15.3$  Hz), 4.21–4.19 (m, 2H), 3.74 (s, 1H), 3.57 (m, 1H), 3.44–3.04 (m, 5H).  $^{13}\text{C}$  NMR (100.5 MHz, acetone- $d_6$ ):  $\delta=169.5$ , 137.7, 137.5, 136.2, 130.5, 129.5, 128.9,

128.6, 128.3, 127.9, 127.4, 126.9, 125.4, 124.5, 123.0, 73.1, 71.9, 52.4, 49.6, 48.8, 48.1, 43.9. Anal. Calcd (%) for  $C_{26}H_{29}N_3O_5S$ : C, 63.01; H, 5.90; N, 8.48. Found: C, 63.23; H, 6.23; N, 8.13.

**4.4.7. 3,4,5,6-Tetrahydro-(4S,5S)-2-[2-(3-methoxyphenyl-acetamido)-benzyl]-7-benzyl-4,5-dihydroxy-1,2,7-thiadiazepine 1,1-dioxide (6e).** White solid. LC–MS,  $m/z=526$  [ $M+H^+$ ].  $^1H$  NMR (400 MHz, acetone- $d_6$ ):  $\delta=8.90$  (br s, 1H), 8.02 (dd, 1H,  $J=1.3, 8.2$  Hz), 7.50–7.29 (m, 7H), 7.20–7.12 (m, 2H), 7.03 (m, 1H), 6.98 (dm, 1H,  $J=7.5$  Hz), 6.97 (ddd, 1H,  $J=1.0, 2.6, 8.3$  Hz), 4.76 (d, 1H,  $J=15.7$  Hz), 4.68 (d, 1H,  $J=15.3$  Hz), 4.55 (d, 1H,  $J=15.7$  Hz), 4.43 (d, 1H,  $J=15.3$  Hz), 4.22–4.19 (m, 2H), 3.76 (s, 3H), 3.71 (s, 2H), 3.58 (m, 1H), 3.40–3.06 (m, 5H).  $^{13}C$  NMR (100.5 MHz, acetone- $d_6$ ):  $\delta=170.1, 160.7, 138.3, 138.2, 138.1, 131.1, 130.2, 129.6, 129.5, 128.9, 128.6, 125.3, 123.9, 122.4, 115.7, 113.1, 73.7, 72.6, 55.4, 53.1, 50.3, 49.4, 48.8, 44.7$ .

**4.4.8. 3,4,5,6-Tetrahydro-(4S,5S)-2-{2-[2-(2-naphthyl)-acetamido]-benzyl}-7-benzyl-4,5-dihydroxy-1,2,7-thiadiazepine 1,1-dioxide (6f).** White solid. LC–MS,  $m/z=546$  [ $M+H^+$ ].  $^1H$  NMR (400 MHz, acetone- $d_6$ ):  $\delta=8.98$  (s, 1H), 8.27 (dm, 1H,  $J=8.4$  Hz), 7.90 (m, 2H), 7.82 (dm, 1H,  $J=8.2$  Hz), 7.61–7.27 (m, 11H), 7.15 (ddd, 1H,  $J=1.2, 7.5, 7.5$  Hz), 4.75 (d, 1H,  $J=15.8$  Hz), 4.66 (d, 1H,  $J=15.4$  Hz), 4.55 (d, 1H,  $J=15.8$  Hz), 4.46 (d, 1H,  $J=15.4$  Hz), 4.27 (s, 1H), 4.26 (d, 1H,  $J=4.4$  Hz), 4.24 (d, 1H,  $J=4.2$  Hz), 3.60 (m, 1H), 3.40–3.10 (m, 5H).  $^{13}C$  NMR (100.5 MHz, acetone- $d_6$ ):  $\delta=169.5, 137.6, 137.5, 134.2, 132.8, 132.5, 130.2, 128.9, 128.74, 128.73, 128.4, 128.2, 127.9, 127.8, 126.4, 125.9, 125.8, 124.8, 124.7, 123.7, 72.9, 72.0, 52.5, 49.5, 48.7, 48.1, 41.4$ . Anal. Calcd (%) for  $C_{30}H_{31}N_3O_5S$ : C, 66.04; H, 5.73; N, 7.70. Found: C, 66.05; H, 5.85; N, 7.68.

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